

ESSENTIAL OILS AND EUGENOL AS ANESTHETICS FOR *Serrasalmus rhombeus**

Ana Paula Gottlieb ALMEIDA¹

Berta Maria HEINZMANN²

Adalberto Luis VAL³

Bernardo BALDISSEROTTO⁴

¹Universidade Federal de Santa Maria – UFSM, Programa de Pós-graduação em Biodiversidade Animal, Avenida Roraima, nº 1000, CEP 97105-900, Santa Maria, RS, Brazil.

²Universidade Federal de Santa Maria – UFSM, Departamento de Farmácia Industrial, Avenida Roraima, nº 1000, CEP 97105-900, Santa Maria, RS, Brazil.

³Instituto Nacional de Pesquisas da Amazônia – INPA, Laboratório de Ecofisiologia e Evolução Molecular, Av. André Araújo, nº 2936, Petrópolis, CEP 69067-375, Manaus, AM, Brazil.

⁴Universidade Federal de Santa Maria – UFSM, Departamento de Fisiologia e Farmacologia, Avenida Roraima, nº 1000, CEP 97105-900, Santa Maria, RS, Brazil. E-mail: bbaldisserotto@hotmail.com (corresponding author).

*Financial support: INCT-ADAPTA (Conselho Nacional de Desenvolvimento Científico e Tecnológico/Fundação de Amparo à Pesquisa no Estado do Amazonas), CNPq, CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior).

Received: March 07, 2017

Approved: October 05, 2017

ABSTRACT

This study evaluated the periods of time of anesthetic induction and recovery of *Serrasalmus rhombeus* exposed to essential oils (EOs) of *Aloysia triphylla* and *Lippia alba* and eugenol, as well as if these anesthetics can be used for transport of this species through analysis of swimming behavior. Fish were placed in aquaria containing different concentrations of *A. triphylla* EO or *L. alba* EO or eugenol, posteriorly were transferred to aquaria containing only water to evaluate the recovery time. In the second experiment, behavior was analyzed during exposure to *A. triphylla* EO, *L. alba* EO or eugenol at 5 or 10 $\mu\text{L L}^{-1}$. The evaluations were carried out at 0, 1, 5, 10 and 15 min of exposure. Fish exposed to 150, 200 and 50 $\mu\text{L L}^{-1}$ of *A. triphylla* EO, *L. alba* EO and eugenol, respectively, showed anesthetic induction time lower than 3 min and recovery time lower than 10 min. Concentrations of 50 $\mu\text{L L}^{-1}$ of both EOs and 25 $\mu\text{L L}^{-1}$ eugenol caused only sedation. Exposure to 5 and 10 $\mu\text{L L}^{-1}$ EOs and eugenol decreased fish swimming time. Both EOs and eugenol were effective for anesthesia and can be used for transport of *S. rhombeus*.

Key words: anesthesia; behavior; black piranha; fish transport; Negro river.

ÓLEOS ESSENCIAIS E EUGENOL COMO ANESTÉSICO PARA *Serrasalmus rhombeus*

RESUMO

Esse estudo avaliou os períodos de tempo de indução e recuperação anestésica de *Serrasalmus rhombeus* expostos aos óleos essenciais (OEs) de *Aloysia triphylla* e *Lippia alba* e eugenol, bem como se esses anestésicos podem ser usados no transporte dessa espécie através da análise do comportamento natatório. Os peixes foram colocados em aquários contendo diferentes concentrações de OE de *A. triphylla* ou OE *L. alba* ou eugenol, posteriormente foram transferidos para aquários contendo somente água para avaliar o tempo de recuperação. No segundo experimento, comportamento foi analisado durante exposição aos OEs ou eugenol nas concentrações 5 ou 10 $\mu\text{L L}^{-1}$. As avaliações foram realizadas em 0, 1, 5, 10 e 15 min de exposição. Peixes expostos a 150, 200 e 50 $\mu\text{L L}^{-1}$ de OE de *A. triphylla*, OE de *L. alba* e eugenol, respectivamente, apresentaram tempo de indução anestésica menor que 3 min e tempo de recuperação menor que 10 min. Concentrações de 50 $\mu\text{L L}^{-1}$ de ambos OEs e 25 $\mu\text{L L}^{-1}$ de eugenol causaram somente sedação. Exposição a 5 e 10 $\mu\text{L L}^{-1}$ de OEs e eugenol diminuíram o tempo de natação dos peixes. Ambos os OEs e o eugenol foram efetivos para anestesia e podem ser utilizados para transporte de *S. rhombeus*.

Palavras-chave: anestesia; comportamento; piranha preta; transporte de peixes; rio Negro.

INTRODUCTION

There is a growing interest in the search for natural anesthetics, obtained from plants, which are economically viable and present low toxicity. The most widely used natural anesthetic is eugenol (4-allyl-2-methoxyphenol), the main compound from clove oil, which is obtained from plants of the genus *Eugenia* (ANDERSON *et al.*, 1997). The anesthetic efficacy of eugenol was showed for several Neotropical species as *Piaractus mesopotamicus* (Holmberg, 1887) (GONÇALVES *et al.*, 2008), *Astyanax bimaculatus* (Linnaeus, 1758) (SILVA *et al.*, 2009), *Brycon amazonicus* (Spix & Agassiz, 1829) (VIDAL *et al.*, 2007), *Rhamdia quelen* (Quoy & Gaimard, 1824) (CUNHA *et al.*, 2010a), *Carassius auratus* (Linnaeus, 1758) (BITTENCOURT *et al.*, 2012), *Centropomus parallelus* Poey, 1860 (SOUZA *et al.*, 2012) and *Brycon hilarii* (Valenciennes, 1850)

(FABIANI *et al.*, 2013). The efficacy of the essential oil (EO) of *Lippia alba* was also demonstrated for *Rhamdia quelen* (CUNHA *et al.*, 2010b; HELDWEIN *et al.*, 2012, 2014; TONI *et al.*, 2014), *Hippocampus reidi* (Ginsburg, 1933) (CUNHA *et al.*, 2011) and *Sparus aurata* (Linnaeus, 1758) (TONI *et al.*, 2015), but the EO of *Aloysia triphylla* was studied only in *R. quelen* (PARODI *et al.*, 2014) and *C. parallelus* (PARODI *et al.*, 2016).

Serrasalmus rhombeus (Linnaeus, 1766), popularly known as black piranha or redeye piranha, is distributed across Amazon and Orinoco basins, Guiana Shield rivers and coastal rivers of northeastern Brazil (JÉGU and INGENITO, 2007). This species is an important fisheries resource (MPA, 2011) and has been exported as ornamental fish (ANJOS *et al.*, 2009), but has powerful teeth that can cause serious injury in humans (MOL, 2006). Therefore, the use of anesthesia in the management of this fish is indicated to prevent injuries in the handlers and to reduce the effects of animal stress.

The aim of this study was to evaluate the anesthetic efficacy of EOs of *A. triphylla* and *L. alba* and eugenol in *S. rhombeus*. In addition, we also investigated the swimming behavior of *S. rhombeus* exposed to low concentrations of these anesthetics to indicate if they could be used in the water of transport of this species.

METHODS

Essential oils and eugenol

The plants *A. triphylla* and *L. alba* were cultivated at the campus of the Universidade Federal de Santa Maria in the city of Frederico Westphalen, southern Brazil. Voucher specimens (SMDB No. 11169 and 10050, respectively) were deposited in the herbarium of the Biology Department. Eugenol (99.9%, Maquira®) was purchased in a local drugstore.

Essential oil extraction and analysis

The oil extraction from the leaves of these plants was performed by hydro-distillation using Clevenger apparatus according to the European Pharmacopoeia (2007). Analysis was made by gas chromatography using an Agilent 7890A gas equipment coupled to an Agilent 5975C mass selective detector (GC-MS). The unit was equipped with a capillary column HP5-MS (Hewlett Packard, 5% fenilmetilsiloxane, 30 m x 0.25 mm, film thickness: 0.25 µm), and the ionization energy used was 70 eV. The parameters chosen for the analysis were: He as gas carrier; split inlet 1:100; temperature program: 40°C for 4 minutes; 40 to 320°C at 4°C min⁻¹; 1 mL min⁻¹ of flow rate; and temperatures of injection and detection of 250°C. The chemical compounds identification was made by comparison of retention indexes, obtained by using a calibration curve of n-alkanes injected at the conditions mentioned for the samples, and the mass fragmentation patterns with NIST (2010) data.

Animals

Specimens of *S. rhombeus* (14.9 ± 0.51 cm, 110.9 ± 3.79 g, voucher number: INPA-ICT 53086) were collected during an expedition to Anavilhanas Islands of the Negro River, 110 km

upstream from Manaus (2°23'41"S, 60°55'14"W). Fish were maintained in tanks with 50% water daily renewed, pumped directly from Rio Negro (29.8°C, pH 5.0), continuously aerated for a few hours before testing.

Experiment I: anesthesia induction and recovery in *S. rhombeus* exposed to *A. triphylla* and *L. alba* EOs and eugenol

The fish were transferred individually to aquaria containing 2 L (29.8 ± 0.46°C; pH 5.0 ± 0.1) of water with the *L. alba* (50, 100 and 200 µL L⁻¹) or *A. triphylla* EOs (50, 100 and 150 µL L⁻¹) or eugenol (25, 40 and 50 µL L⁻¹), first diluted in ethanol at a proportion of 1:10. These concentrations were chosen based in previous studies (CUNHA *et al.*, 2010a, b; PARODI *et al.*, 2014). A total of 14 compounds were identified in each essential oil. The main constituents of *A. triphylla* EO were limonene (21.69%) and geranial (24.32%) and of *L. alba* EO were linalool (66.35%) and eucalyptol (10.63%) (Table 1).

The time for anesthesia induction was evaluated according to SMALL (2003): stage I - corresponds to sedation, when the reactivity to external stimuli decreased; stage II - corresponds to partial loss of equilibrium and erratic swimming; and stage III - corresponds to total loss of equilibrium and cessation of locomotion. In recovery, the fish returns to regular swimming. Eight fish were used for each tested concentration and each fish was used only once. The maximum observation time was 15 min, since several studies indicated that sedation and anesthesia occur within this period (CÁRDENAS *et al.*, 2016; HOHLENWERGER *et al.*, 2016; PARODI *et al.*, 2016; SENA *et al.*, 2016; TEIXEIRA *et al.*, 2017). Control experiment was performed using aquaria containing water and ethanol at a concentration equivalent to the highest dilution (1800 µL L⁻¹). After induction of anesthesia, fish were transferred to a tank containing only water to evaluate the recovery time. The animals were recovered when swimming regularly and reacting to external stimuli (the peduncle of the caudal fin was pressed with a glass rod).

Experiment II: fish behavior through exposure to low concentrations of *A. triphylla* and *L. alba* EOs and eugenol

The fish were placed into tanks containing 20 L of water and *A. triphylla* or *L. alba* EOs or eugenol at 5 and 10 µL L⁻¹. These concentrations were 2.5 and 5-fold lower than the lowest eugenol concentration tested, which induced stage II (partial loss of equilibrium) (see results) and also based on studies carried out by BECKER *et al.* (2012, 2013) and PARODI *et al.* (2014). Four fish per aquaria were used for each concentration (in triplicate). The fish were filmed for 20 s for analysis of the total swimming time and equilibrium (partial or total loss of equilibrium or normal) at 0, 1, 5, 10 and 15 min of exposure. Control experiments were performed using aquaria containing only water and aquaria containing ethanol at a concentration equivalent to that used in the highest dilution of the EOs (90 µL L⁻¹).

Table 1. Chemical composition of essential oils.

Essential Oil	RI* Experimental	RI Literature ^a	Chemical Compound	Percent Composition
<i>A. triphylla</i>	989	986	5-Hepten-2-one, 6-methyl	2.085
	991	986	β -Pinene	0.499
	1026	1026	Limonene	21.694
	1049	1048	β -cis-Ocimene	0.717
	1229	1228	cis-Geraniol	2.218
	1241	1247	cis-Carveol	18.533
	1256	1259	Linlyl Acetate	2.703
	1271	1269	Geranial	24.317
	1417	1415	β -Caryophyllene	5.323
	1483	1483	α -Curcumene	3.251
	1495	1487	(-)-Alloaromadendrene	1.136
	1577	1578	Spathulenol	2.617
	1582	1583	Caryophyllene Oxide	6.793
	1640	1639	T-Cadinol	1.411
			Identified compounds	93.297
<i>L. alba</i>	971	969	Sabinene	0.817
	992	996	β -Pinene	0.972
	1027	1026	Limonene	1.992
	1028	1030	Eucalyptol	10.633
	1100	1101	Linalool	66.347
	1143	1146	Camphor	0.516
	1204	1205	Trans-Dihydrocarvone	1.183
	1242	1252	Carveol	1.135
	1272	1270	Geranial	0.764
	1273	1270	Neral	0.361
	1418	1419	Aromadendrene	3.480
	1480	1482	Germacrene D	2.784
	1556	1558	Germacrene B	2.219
	1582	1582	Spathulenol	1.340
			Identified Compounds	94.543

*RI = Retention Index. ^aNIST, 2010.

Statistical analyses

All data are expressed as mean \pm SEM. Homogeneity of variances among treatments was tested by Levene's test. The data from time to induction of anesthetic stages presented homogeneous variances and comparisons between the different concentrations were assessed using one-way ANOVA and Tukey's test. The data from swimming time did not exhibit homogeneous variances; therefore, comparisons between the different treatments and times were assessed using the non-parametric Scheirer-Ray-Hare extension of the Kruskal-Wallis test followed by the post-hoc Nemenyi test. The analysis was performed using the Statistica 7.0 software (Stat Soft. Inc.) and the minimum significance level was set at $P < 0.05$.

RESULTS

The concentrations of 50 $\mu\text{L L}^{-1}$ *A. triphylla* EO, 50 $\mu\text{L L}^{-1}$ *L. alba* EO and 25 $\mu\text{L L}^{-1}$ eugenol only induced sedative effect within 15 min, but all other assay concentrations induced all stages of anesthesia. Fish exposed to 150, 200 and 50 $\mu\text{L L}^{-1}$ of *A. triphylla* and *L. alba* EO and eugenol, respectively, reached deep anesthesia (stage 3) significantly faster than the lower concentrations (Figure 1). The increasing concentration of *L. alba* EO increased proportionally recovery time (Figure 1B). The *A. triphylla* EO increased recovery time up to 100 $\mu\text{L L}^{-1}$ and eugenol only at the highest concentration (50 $\mu\text{L L}^{-1}$) (Figures 1A, C). The fish showed at the first contact with the anesthetics in both tests small jumps and swimming bursts in the first experiment. The application of 1800 $\mu\text{L L}^{-1}$ ethanol alone did not produce any anesthetic effect.

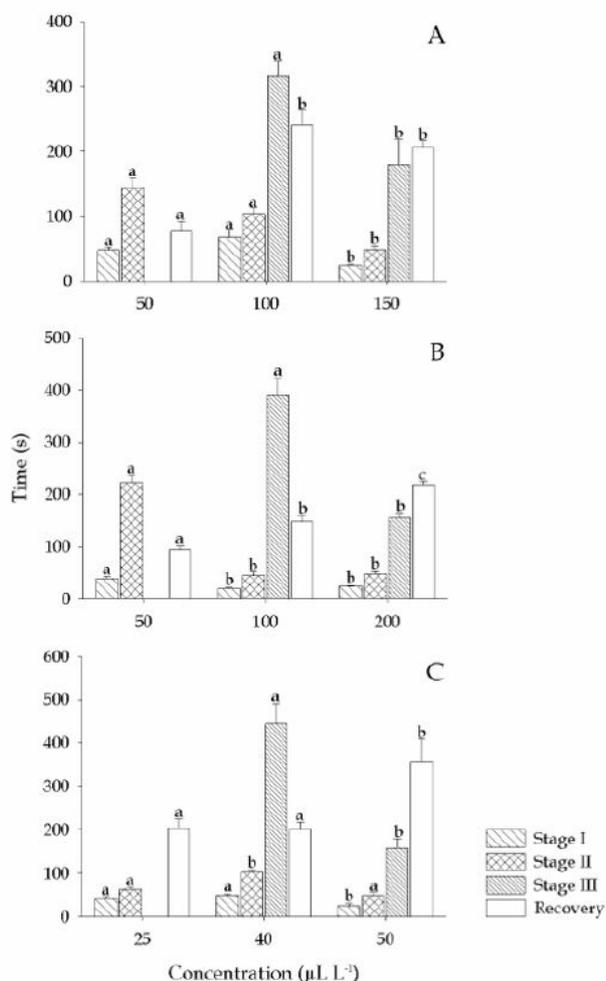


Figure 1. Time required for induction and recovery of anesthesia in *Serrasalmus rhombeus* using *Aloysia triphylla* (A) and *Lippia alba* (B) essential oils and eugenol (C). Different letters indicate a significant difference in the same stage based on one-way ANOVA and Tukey's test ($P < 0.05$).

In the second experiment, fish exposed to 10 $\mu\text{L L}^{-1}$ EO of *A. triphylla*, 5 and 10 $\mu\text{L L}^{-1}$ EO of *L. alba* and 5 $\mu\text{L L}^{-1}$ of eugenol presented lower swimming time than control fish in all evaluated times. The fish did not show loss of equilibrium at both concentrations of EOs and eugenol tested. Ethanol-exposed fish initially decreased swimming time, but returned to the control level after 10 min (Figure 2).

DISCUSSION

Aloysia triphylla EO used in this study presented in its chemical composition limonene and geranial as main constituents. Unlike the present study, other studies detected citral, which is composed of geranial and neral, as main constituents of this EO (FIGUEIREDO *et al.*, 2004;

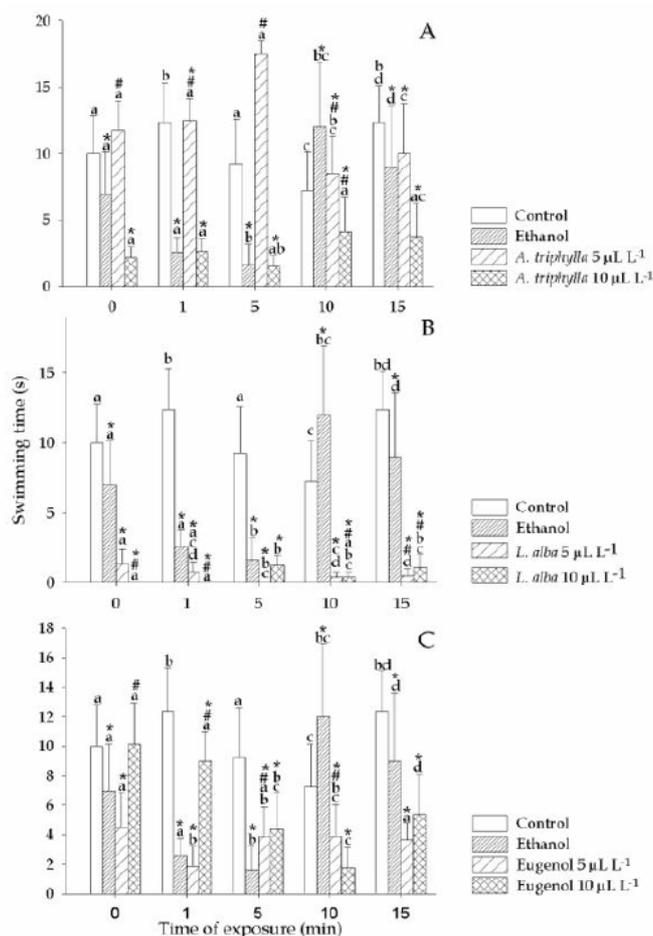


Figure 2. Swimming time of *Serrasalmus rhombeus* exposed to low concentrations of *Aloysia triphylla* (A) and *Lippia alba* (B) essential oils and eugenol (C). Different letters indicate significant difference between times of exposure. *indicates significant difference from control fish. # indicates significant difference from ethanol-exposed fish ($P < 0.05$).

SARTORATTO *et al.*, 2004; PAULUS *et al.*, 2013). *Lippia alba* EO presented as the main constituent linalool, as observed in previous studies (HELDWEIN *et al.*, 2012; TONI *et al.*, 2015).

The anesthetic and sedative effects of eugenol are well established in rodents and fish (CUNHA *et al.*, 2010a; FREIRE *et al.*, 2006). On the other hand, the central depressor effects detected for the EOs of *A. triphylla* and *L. alba* are due to the association of different components, and resulted from additive and/or synergistic activities. For some of their constituents, as linalool and spathulenol, anesthetic and sedative effects were already described in *Rhamdia quelen* (BENOVIT *et al.*, 2015; HELDWEIN *et al.*, 2014). Additionally, some components detected in the EOs showed sedative and/ or anxiolytic like properties in mice, as limonene, geranial, 1,8-cineole and β -pinene

(GOMES *et al.*, 2010; GUZMÁN-GUTIÉRREZ *et al.*, 2012; VALE *et al.*, 2002).

The present study demonstrates that eugenol and the EOs of *A. triphylla* and *L. alba* presented anesthetic effect in *S. rhombeus*. According to PARK *et al.* (2009), the suitable anesthetic induction time is about 3 min and at most 10 minutes for recovery. Following this premise, the best concentrations of the EOs of *A. triphylla* and *L. alba* and eugenol to anesthetize *S. rhombeus* were 150, 200 and 50 $\mu\text{L L}^{-1}$, respectively. The time required to induce deep anesthesia at these concentrations was approximately 164 s. In relation to the recovery time, all concentrations of EOs and eugenol remained in the suggested range by PARK *et al.* (2009), the largest time being 355 s. The concentration of the EO of *A. triphylla* to anesthetize both albino and gray strains of *R. quelen* at 24°C and pH 7.0 within 3 min was between 400-800 $\mu\text{L L}^{-1}$ (PARODI *et al.*, 2014). The lowest concentration of the EO of *L. alba* that induced anesthesia in *R. quelen* within 3 min at 21°C and pH 7.0 was 400 mg L^{-1} (around 500 $\mu\text{L L}^{-1}$ because the density of this EO is of approximately 0.8) (CUNHA *et al.*, 2010a), and in *H. reidi* was 450 $\mu\text{L L}^{-1}$ (CUNHA *et al.*, 2011). Therefore, for both EOs deep anesthesia can be obtained with lower concentration in *S. rhombeus* than in these other species studied, probably due to the higher temperature used in the present study as found by GOMES *et al.* (2011). In contrast, *S. rhombeus* exposed to 50 $\mu\text{L L}^{-1}$ did not reach anesthesia stage, but *H. reidi* did (CUNHA *et al.*, 2011). TONI *et al.* (2015) observed that the lowest concentration required to induce deep anesthesia in *Sparus aurata* at 38 ppt salinity, 18°C, within 3 min was 200 $\mu\text{L L}^{-1}$ EO of *L. alba*, the same concentration found to *S. rhombeus*. As for *S. rhombeus*, all concentrations tested in *S. aurata* showed recovery time lower than 10 min (TONI *et al.*, 2015).

The lowest eugenol concentration necessary to anesthetize *S. rhombeus* within 3 min was 50 $\mu\text{L L}^{-1}$. This is the same lowest eugenol concentration to anesthetize *R. quelen* (at 21°C and pH 7) (CUNHA *et al.*, 2010b), *B. amazonicus* (*B. cephalus*) (VIDAL *et al.*, 2007) and *P. mesopotamicus* (at 25 °C) (GONÇALVES *et al.*, 2008). *Centropomus parallelus* needed a similar eugenol concentration range to anesthetize and recover within the proposed periods: 25 – 62.5 mg L^{-1} (23.6 – 59 $\mu\text{L L}^{-1}$, because the density of eugenol is approximately 1.06) at 21°C (SOUZA *et al.*, 2012). *Brycon hilarii* needed higher concentrations than *S. rhombeus*, in the range of 100 – 300 mg L^{-1} (94.3 – 283 $\mu\text{L L}^{-1}$), for induction and recovery at the suitable time at 25°C (FABIANI *et al.*, 2013). In contrast, BITTENCOURT *et al.* (2012) verified that 75 mg L^{-1} (70.7 $\mu\text{L L}^{-1}$) eugenol required more than 3 min to anesthetize *C. auratus* (higher concentrations were not tested). The 5.0-7.0 pH range at 23°C did not change time of induction to eugenol in *R. quelen* exposed to 40 mg L^{-1} (37.7 $\mu\text{L L}^{-1}$), but at pH 7.0 and 30°C eugenol anesthetized this species within 225-275 s (depending on size of the fish) (GOMES *et al.*, 2011). *S. rhombeus* needs a higher time (443 s) to anesthetize at this eugenol concentration.

In the second experiment, the exposure of the fish to low concentrations of the anesthetics was performed to verify

the possibility of using these products in the transport of *S. rhombeus*. The sedation stage, with lower responsiveness to external stimuli and metabolic rate, but without losing equilibrium, is recommended for transporting fish (SUMMERFELT and SMITH, 1990; PIRHONEN and SCHRECK, 2003).

Ethanol initially reduced swimming activity, but it returned to control level after 10 min. Ethanol enhances the action of several GABA receptors subtypes (WALLNER *et al.*, 2003), which may be related to the decreased swimming activity in *S. rhombeus*. This effect may have been fast due to ethanol evaporation. Both concentrations of EO of *A. triphylla* can be tested for transport of *S. rhombeus*, and the best concentration apparently is 10 $\mu\text{L L}^{-1}$, which reduced swimming activity through the 15 min observation. The addition of 30 to 50 $\mu\text{L L}^{-1}$ of EO of *A. triphylla* in transport water (lower concentrations were not tested) reduced ions loss, plasma cortisol levels and ammonia excretion in *R. quelen*, suggesting lower physiological damage resulting from transport (PARODI *et al.*, 2014; ZEPPENFELD *et al.*, 2014). Both EO of *L. alba* concentrations can be tested for the transport of *S. rhombeus*. Similar concentrations (10 and 20 $\mu\text{L L}^{-1}$) of *L. alba* EO are recommended for the transport of *R. quelen* because they improved blood and ionoregulatory parameters (BECKER *et al.*, 2012). Both concentrations of eugenol can be tested for the transport of *S. rhombeus*, and the best concentration seems to be 5 $\mu\text{L L}^{-1}$, which reduced swimming activity at all observation times. The addition of 1 – 3 $\mu\text{L L}^{-1}$ of eugenol is recommended for the transport of *R. quelen* because they decreased non-ionized ammonia levels, ion loss and mortality (BECKER *et al.*, 2012, 2013). The determination of the swimming activity allowed a more precise analysis of the sedative effects of the anesthetics tested, because the identification of the decreased reactivity to external stimuli proposed by SMALL (2003) for stage I is rather subjective. However, since this study analyzed swimming activity for only 15 min, it would be interesting that further studies investigate if the effects on swimming activity and equilibrium caused by the essential oils and eugenol can last for several hours, as well as if they can improve water, blood and ionoregulatory parameters as observed in the transport of silver catfish as observed by BECKER *et al.* (2012, 2013), PARODI *et al.* (2014) and ZEPPENFELD *et al.* (2014).

CONCLUSION

In conclusion, both *A. triphylla* and *L. alba* EOs and eugenol are effective for anesthetic induction within 3 min at the concentrations 150, 200 e 50 $\mu\text{L L}^{-1}$, respectively. For fast sedation, the recommended concentration is 50 $\mu\text{L L}^{-1}$ for both EOs and 25 $\mu\text{L L}^{-1}$ for eugenol. The concentrations of 5 and 10 $\mu\text{L L}^{-1}$ of both EOs and eugenol reduced fish swimming activity and are indicated for studies of transportation of *S. rhombeus* but for eugenol and the EO of *A. triphylla* the best concentrations are 5 and 10 $\mu\text{L L}^{-1}$, respectively.

ACKNOWLEDGEMENTS

The authors acknowledge support from INCT-ADAPTA (Conselho Nacional de Desenvolvimento Científico e Tecnológico/Fundação de Amparo à Pesquisa no Estado do Amazonas). A.L. Val, B. Baldisserotto and B.M. Heinzmann received CNPq research fellowships and A.P.G. Almeida received CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior) PhD fellowship.

REFERENCES

- ANDERSON, W.G.; MCKINLEY, R.S.; COLAVECCHIA, M. 1997 The use of clove oil as an anesthetic for rainbow trout and its effects on swimming performance. *North American Journal of Fisheries Management*, 17(2): 301-307. [http://dx.doi.org/10.1577/1548-8675\(1997\)017<0301:TUO COA>2.3.CO;2](http://dx.doi.org/10.1577/1548-8675(1997)017<0301:TUO COA>2.3.CO;2).
- ANJOS, H.D.B.; AMORIM, R.M.S.; SIQUEIRA, J.A.; ANJOS, C.R. 2009 Exportação de peixes ornamentais do estado do Amazonas, bacia Amazônica, Brasil. *Boletim do Instituto de Pesca*, 35(2): 259-274.
- BECKER, A.G.; CUNHA, M.A.; GARCIA, L.O.; ZEPPEFELD, C.C.; PARODI, T.V.; MALDANER, G.; MOREL, A.F.; BALDISSEROTTO, B. 2013 Efficacy of eugenol and the methanolic extract of *Condalia buxifolia* during the transport of the silver catfish *Rhamdia quelen*. *Neotropical Ichthyology*, 11(3): 675-681. <http://dx.doi.org/10.1590/S1679-62252013000300021>.
- BECKER, A.G.; PARODI, T.V.; HELDWEIN, C.G.; ZEPPEFELD, C.C.; HEINZMANN, B.M.; BALDISSEROTTO, B. 2012 Transportation of silver catfish, *Rhamdia quelen*, in water with eugenol and the essential oil of *Lippia alba*. *Fish Physiology and Biochemistry*, 38(3): 789-796. PMID:21972065. <http://dx.doi.org/10.1007/s10695-011-9562-4>.
- BENOVIT, S.C.; SILVA, L.L.; SALBEGO, J.; LORO, V.L.; MALLMANN, C.A.; BALDISSEROTTO, B.; FLORES, E.M.M.; HEINZMANN, B.M. 2015 Anesthetic activity and bio-guided fractionation of the essential oil of *Aloysia gratissima* (Gillies & Hook.) Tronc. in silver catfish *Rhamdia quelen*. *Anais da Academia Brasileira de Ciências*, 87(3): 1675-1689. PMID:26221984. <http://dx.doi.org/10.1590/0001-3765201520140223>.
- BITTENCOURT, F.; SOUZA, B.E.; BOSCOLO, W.R.; RORATO, R.R.; FEIDEN, A.; NEU, D.H. 2012 Benzocaína e eugenol como anestésicos para quinguio (*Carassius auratus*). *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 64(6): 1597-1602. <http://dx.doi.org/10.1590/S0102-09352012000600028>.
- CÁRDENAS, C.; TONI, C.; MARTOS-SITCHA, J.A.; CÁRDENAS, S.; HERAS, V.; BALDISSEROTTO, B.; HEINZMANN, B.M.; VÁZQUEZ, R.; MANCERA, J.M. 2016 Effects of clove oil, essential oil of *Lippia alba* and 2-phe anaesthesia on juvenile meagre, *Argyrosomus regius* (Asso, 1801). *Journal of Applied Ichthyology*, 32(4): 693-700. <http://dx.doi.org/10.1111/jai.13048>.
- CUNHA, M.A.; ZEPPEFELD, C.C.; GARCIA, L.O.; LORO, V.L.; FONSECA, M.B.; EMANUELLI, T.; VEECK, A.P.L.; COPATTI, C.E.; BALDISSEROTTO, B. 2010a Anesthesia of silver catfish with eugenol: time of induction, cortisol response and sensory analysis of fillet. *Ciência Rural*, 40(10): 2107-2114. <http://dx.doi.org/10.1590/S0103-84782010001000009>.
- CUNHA, M.A.; BARROS, F.M.C.; GARCIA, L.O.; VEECK, A.P.L.; HEINZMANN, B.M.; LORO, V.L.; EMANUELLI, T.; BALDISSEROTTO, B. 2010b Essential oil of *Lippia alba*: a new anesthetic for silver catfish, *Rhamdia quelen*. *Aquaculture*, 306(1-4): 403-406. <http://dx.doi.org/10.1016/j.aquaculture.2010.06.014>.
- CUNHA, M.A.; SILVA, B.F.; DELUNARDO, F.A.C.; BENOVI, S.C.; GOMES, L.C.; HEINZMANN, B.M.; BALDISSEROTTO, B. 2011 Anesthetic induction and recovery of *Hippocampus reidi* exposed to the essential oil of *Lippia alba*. *Neotropical Ichthyology*, 9(3): 683-688. <http://dx.doi.org/10.1590/S1679-62252011000300022>.
- EUROPEAN PHARMACOPOEIA. 2007 European directorate for the quality of medicines. 6th ed. Strassbourg: European Pharmacopoeia.
- FABIANI, B.M.; BOSCOLO, W.R.; FEIDEN, A.; DIEMER, O.; BITTENCOURT, F.; NEU, D.H. 2013 Benzocaine and eugenol as anesthetics for *Brycon hilarii*. *Acta Scientiarum Animal Science*, 35(2): 113-117.
- FIGUEIREDO, R.O.; STEFANINI, M.B.; MING, L.C.; MARQUES, M.O.M.; FACANALI, R. 2004 Essential oil composition of *Aloysia triphylla* (L'Herit) Britton leaves cultivated in Botucatu, São Paulo, Brazil. *Acta Horticulturae*, 629(1): 131-134. <http://dx.doi.org/10.17660/ActaHortic.2004.629.17>.
- FREIRE, C.M.M.; MARQUES, M.O.M.; COSTA, M. 2006 Effects of seasonal variation on the central nervous system activity of *Ocimum gratissimum* L. essential oil. *Journal of Ethnopharmacology*, 105(1-2): 161-166. PMID:16303272. <http://dx.doi.org/10.1016/j.jep.2005.10.013>.
- GOMES, D.P.; CHAVES, B.W.; BECKER, A.G.; BALDISSEROTTO, B. 2011 Water parameters affect anaesthesia induced by eugenol in silver catfish, *Rhamdia quelen*. *Aquaculture Research*, 42(6): 878-886. <http://dx.doi.org/10.1111/j.1365-2109.2011.02864.x>.
- GOMES, P.B.; FEITOSA, M.L.; SILVA, M.I.; NORONHA, E.C.; MOURA, B.A.; VENÂNCIO, E.T.; RIOS, E.R.; SOUSA, D.P.; VASCONCELOS, S.M.; FONTELES, M.M.; SOUSA, F.C. 2010 Anxiolytic-like effect of the monoterpene 1,4-cineole in mice. *Pharmacology, Biochemistry, and Behavior*, 96(3): 287-293. PMID:20670917. <http://dx.doi.org/10.1016/j.pbb.2010.05.019>.
- GONÇALVES, A.F.N.; SANTOS, E.C.C.; FERNANDES, J.B.K.; TAKAHASHI, L.S. 2008 Mentol e eugenol como substitutos da benzocaína na indução anestésica de juvenis de pacu. *Acta Scientiarum. Animal Science (Penicuik, Scotland)*, 30(3): 339-344.
- GUZMÁN-GUTIÉRREZ, S.L.; GÓMEZ-CANSINO, R.; GARCÍA-ZEBADÚA, J.C.; JIMÉNEZ-PÉREZ, N.C.; REYES-CHILPA, R. 2012 Antidepressant activity of *Litsea glaucescens* essential oil: Identification of β -pinene and linalool as active principles. *Journal of Ethnopharmacology*, 143(2): 673-679. PMID:22867633. <http://dx.doi.org/10.1016/j.jep.2012.07.026>.
- HELDWEIN, C.G.; SILVA, L.L.; GAI, E.Z.; ROMAN, C.; PARODI, T.V.; BÜRGER, M.E.; BALDISSEROTTO, B.; FLORES, E.M.M.; HEINZMANN, B.M. 2014 S-(+)-Linalool from *Lippia alba*: sedative and anesthetic for silver catfish (*Rhamdia quelen*). *Veterinary Anaesthesia and Analgesia*, 41(6): 621-629. PMID:24628858. <http://dx.doi.org/10.1111/vaa.12146>.
- HELDWEIN, C.G.; SILVA, L.L.; RECKZIEGEL, P.; BARROS, F.M.C.; BÜRGER, M.E.; BALDISSEROTTO, B.; MALLMANN, C.A.; SCHMIDT, D.; CARON, B.O.; HEINZMANN, B.M. 2012 Participation of the GABAergic system in the anesthetic effect of *Lippia alba* (Mill.) N.E. Brown essential oil. *Brazilian Journal of Medical and Biological Research*, 45(5): 436-443. PMID:22473320. <http://dx.doi.org/10.1590/S0100-879X2012007500052>.

- HOHLENWERGER, J.C.; COPATTI, C.E.; SENA, A.C.; COUTO, R.D.; BALDISSEROTTO, B.; HEINZMANN, B.M.; CARON, B.O.; SCHMIDT, D. 2016 Could the essential oil of *Lippia alba* provide a readily available and cost-effective anaesthetic for Nile tilapia (*Oreochromis niloticus*)? *Marine and Freshwater Behaviour and Physiology*, 49(2): 119-126. <http://dx.doi.org/10.1080/10236244.2015.1123869>.
- JÉGU, M.; INGENITO, L.F.S. 2007 Serrasalminae. In: BUCKUP, P.A.; MENEZES, N.A.; GHAZZI, M.S. *Catálogo das espécies de peixes de água doce do Brasil*. Rio de Janeiro: Museu Nacional. p. 40-43.
- MOL, J.H. 2006 Attacks on humans by the piranha *Serrasalmus rhombeus* in Suriname. *Studies on Neotropical Fauna and Environment*, 41(3): 189-195. <http://dx.doi.org/10.1080/01650520600630683>.
- MPA – Ministério da Pesca e Aquicultura. 2011 *Boletim estatístico da pesca e aquicultura*. Brasília: MPA. 59 p.
- NIST; EPA; NIH. 2010 *Mass spectral library and search/ analysis programs*. Hoboken: J. Wiley and Sons.
- PARK, I.S.; PARK, M.O.; HUR, J.M.; KIM, D.S.; CHANG, Y.J.; KIM, Y.J.; PARK, J.Y.; JOHNSON, S.C. 2009 Anesthetic effects of lidocaine hydrochloride on water parameters in simulated transport experiment of juvenile winter flounder *Pleuronectes americanus*. *Aquaculture*, 294(1-2): 76-79. <http://dx.doi.org/10.1016/j.aquaculture.2009.05.011>.
- PARODI, T.V.; CUNHA, M.A.; BECKER, A.G.; ZEPPENFELD, C.C.; MARTINS, D.I.; KOAKOSKI, G.; BARCELLOS, L.G.; HEINZMANN, B.M.; BALDISSEROTTO, B. 2014 Anesthetic activity of the essential oil of *Aloysia triphylla* and effectiveness in reducing stress during transport of albino and gray strains of silver catfish, *Rhamdia quelen*. *Fish Physiology and Biochemistry*, 40(2): 323-334. PMID:23974669. <http://dx.doi.org/10.1007/s10695-013-9845-z>.
- PARODI, T.V.; SANTOS, C.A.; VERONEZ, A.; GOMES, L.C.; HEINZMANN, B.M.; BALDISSEROTTO, B. 2016 Anesthetic induction and recovery time of *Centropomus parallelus* exposed to the essential oil of *Aloysia triphylla*. *Ciência Rural*, 46(12): 2142-2147. <http://dx.doi.org/10.1590/0103-8478cr20160039>.
- PAULUS, D.; VALMORBIDA, R.; TOFFOLI, E.; NAVA, G.A.; PAULUS, E. 2013 Teor e composição química do óleo essencial e crescimento vegetativo de *Aloysia triphylla* em diferentes espaçamentos e épocas de colheita. *Revista Ceres*, 60(3): 372-379. <http://dx.doi.org/10.1590/S0034-737X2013000300010>.
- PIRHONEN, J.; SCHRECK, C.B. 2003 Effects of anesthesia with MS-222, clove oil and CO₂ on feed intake and plasma cortisol in steelhead trout (*Oncorhynchus mykiss*). *Aquaculture (Amsterdam, Netherlands)*, 220(1-4): 507-514. [http://dx.doi.org/10.1016/S0044-8486\(02\)00624-5](http://dx.doi.org/10.1016/S0044-8486(02)00624-5).
- SARTORATTO, A.; MACHADO, A.L.M.; DELARMELENA, C.; FIGUEIRA, G.M.; DUARTE, M.C.T.; REHDER, V.L.G. 2004 Composition and antimicrobial activity of essential oils from aromatic plants used in Brazil. *Brazilian Journal of Microbiology*, 35(4): 275-280. <http://dx.doi.org/10.1590/S1517-83822004000300001>.
- SENA, A.C.; TEIXEIRA, R.R.; FERREIRA, E.L.; HEINZMANN, B.M.; BALDISSEROTTO, B.; CARON, B.O.; SCHMIDT, D.; COUTO, R.D.; COPATTI, C.E. 2016 Essential oil from *Lippia alba* has anaesthetic activity and is effective in reducing handling and transport stress in tambacu (*Piaractus mesopotamicus* × *Colossoma macropomum*). *Aquaculture (Amsterdam, Netherlands)*, 465(1): 374-379. <http://dx.doi.org/10.1016/j.aquaculture.2016.09.033>.
- SILVA, E.M.P.; OLIVEIRA, R.H.F.; RIBEIRO, M.A.R.; COPPOLA, M.P. 2009 Efeito anestésico do óleo de cravo em alevinos de lambari. *Ciência Rural*, 39(6): 1851-1856. <http://dx.doi.org/10.1590/S0103-84782009005000127>.
- SMALL, B.C. 2003 Anesthetic efficacy of metomidate and comparison of plasma cortisol responses to tricaine methanesulfonate, quinaldine and clove oil anesthetized channel catfish *Ictalurus punctatus*. *Aquaculture (Amsterdam, Netherlands)*, 218(1-4): 177-185. [http://dx.doi.org/10.1016/S0044-8486\(02\)00302-2](http://dx.doi.org/10.1016/S0044-8486(02)00302-2).
- SOUZA, R.A.R.; CARVALHO, C.V.A.; NUNES, F.F.; SCOPEL, B.R.; GUARIZI, J.D.; TSUZUKI, M.Y. 2012 Efeito comparativo da benzocaina, mentol e eugenol como anestésicos para juvenis de robalo peva. *Boletim do Instituto de Pesca*, 38(3): 247-255.
- SUMMERFELT, R.C.; SMITH, L.S. 1990 Anaesthesia, surgery, and related techniques. In: SCHRECK, C.B.; MOYLE, P.B. *Methods for fish biology*. Bethesda: American Fisheries Society. p. 213-272.
- TEIXEIRA, R.R.; SOUZA, R.C.; SENA, A.C.; BALDISSEROTTO, B.; HEINZMANN, B.M.; COUTO, R.D.; COPATTI, C.E. 2017 Essential oil of *Aloysia triphylla* in Nile tilapia: anaesthesia, stress parameters and sensory evaluation of fillets. *Aquaculture Research*, 48(7): 3383-3392. <http://dx.doi.org/10.1111/arc.13165>.
- TONI, C.; BECKER, A.G.; SIMÕES, L.N.; PINHEIRO, C.G.; SILVA, L.L.; HEINZMANN, B.M.; CARON, B.O.; BALDISSEROTTO, B. 2014 Fish anesthesia: effects of the essential oils of *Hesperozygis ringens* and *Lippia alba* on the biochemistry and physiology of silver catfish (*Rhamdia quelen*). *Fish Physiology and Biochemistry*, 40(3): 701-714. PMID:24141557. <http://dx.doi.org/10.1007/s10695-013-9877-4>.
- TONI, C.; MARTOS-SITCHA, J.A.; BALDISSEROTTO, B.; HEINZMANN, B.M.; SILVA, L.L.; MARTÍNEZ-RODRÍGUEZ, G.; MANCERA, J.M. 2015 Sedative effect of 2-phenoxyethanol and essential oil of *Lippia alba* on stress response in gilthead sea bream (*Sparus aurata*). *Research in Veterinary Science*, 103(1): 20-27. PMID:26679791. <http://dx.doi.org/10.1016/j.rvsc.2015.09.006>.
- VALE, T.G.; FURTADO, E.C.; SANTOS JUNIOR, J.G.; VIANA, G.S. 2002 Central effects of citral, myrcene and limonene, constituents of essential oil chemotypes from *Lippia alba* (Mill.) N.E. Brown. *Phytomedicine*, 9(8): 709-714. PMID:12587690. <http://dx.doi.org/10.1078/094471102321621304>.
- VIDAL, L.V.O.; FURUYA, W.M.; GRACIANO, T.S.; SCHAMBER, C.R.; SILVA, L.C.R.; SANTOS, L.D.; SOUZA, S.R. 2007 Eugenol como anestésico para juvenis de matrinxã (*Brycon cephalus*). *Revista Brasileira de Saúde e Produção Animal*, 8(4): 335-342.
- WALLNER, M.; HANCHAR, H.J.; OLSEN, R.W. 2003 Ethanol enhances 4,3 and 6,3-aminobutyric acid type A receptors at low concentrations known to affect humans. *Proceedings of the National Academy of Sciences of the United States of America*, 100(25): 15218-15223. PMID:14625373. <http://dx.doi.org/10.1073/pnas.2435171100>.
- ZEPPENFELD, C.C.; TONI, C.; BECKER, A.G.; MIRON, D.S.; PARODI, T.V.; HEINZMANN, B.M.; BARCELLOS, L.J.G.; KOAKOSKI, G.; ROSA, J.G.S.; LORO, V.L.; CUNHA, M.A.; BALDISSEROTTO, B. 2014 Physiological and biochemical responses of silver catfish, *Rhamdia quelen*, after transport in water with essential oil of *Aloysia triphylla* (L'Herit) Britton. *Aquaculture*, 418-419(1): 101-107. <http://dx.doi.org/10.1016/j.aquaculture.2013.10.013>.